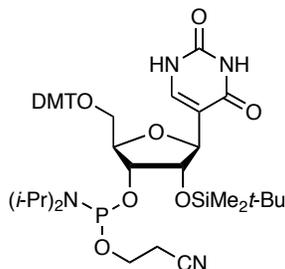


## Pseudouridine CEP (BA 0280)

### Product Information



Pseudouridine ( $\Psi$ ) is one of the most common modified nucleosides found in RNA, e.g., in tRNAs and snRNAs.<sup>1</sup> The uracil nucleobase is identical to that found in uridine except that it is attached to the ribose ring via C5 rather than N1, i.e., it is a C-nucleoside. Thus, in addition to the ability to form Watson-Crick base pairs with adenosine in the normal manner,  $\Psi$  has an additional hydrogen bond donor at N1. This difference can strongly influence the overall structure of an RNA oligonucleotide.<sup>2</sup> As an example of the consequences of the N1 hydrogen, the ability to coordinate a structural water molecule may result in rigidifying the nearby sugar-phosphate backbone and enhancing base stacking.<sup>3</sup>

The ability to install a  $\Psi$  residue site-specifically allows the systematic study of its effect on the structure, function and stability of RNA. Several strategies have been reported for the incorporation of  $\Psi$  during the chemical synthesis of RNA oligonucleotides.<sup>3-7</sup> We now offer Pseudouridine CEP ( $\Psi$  CEP, BA 0280) for this purpose. This particular version<sup>3,6</sup> of pseudouridine phosphoramidite relies on standard cyanoethyl phosphoramidite coupling chemistry, 2'-*O*-TBDMS protection, and no nucleobase protecting groups.<sup>5</sup>

The availability of totally synthetic  $\Psi$ -containing oligoribonucleotides has led to the synthesis of modified ribozymes and key portions of natural tRNAs and snRNAs and has generated numerous observations about the role of  $\Psi$  in RNA.<sup>2-5,7,8,9</sup> For example,<sup>1</sup> in double-stranded RNA, the N1 hydrogen projects into the deep and narrow major groove, and <sup>1</sup>H-NMR studies on synthetic duplex A-form RNA show that the uniquely visible<sup>2,3,8</sup> N1 hydrogen is normally non-bonded, but may be accessed with metal ions, spermidine, and charged peptide side chains. Replacement or addition of pseudouridine residues in synthetic anticodon domains of tRNA<sup>Lys</sup> (human and *E. coli*) had a dramatic effect on its structure.<sup>8</sup> In studies on synthetic fragments of 23S rRNA, altering of the number and position of  $\Psi$  residues showed a range of effects, both stabilizing and destabilizing.<sup>7</sup> It was proposed that  $\Psi$  may be stabilizing relative to U because of greater hydrophilicity, presumably due to additional hydrogen bonding via the N1 hydrogen.<sup>7</sup> It is hoped that the availability of Pseudouridine CEP may facilitate further research on the customization of the structure and function of RNA oligonucleotides.

**Coupling, cleavage, and nucleobase deprotection:** In our hands, standard RNA coupling protocols using a 12 minute coupling time were successful in achieving >95%

coupling yields. Cleavage and nucleobase protection was accomplished by first passing 3:1 concentrated ammonium hydroxide:ethanol through the column over 1-1.5 h, then heating the eluate overnight at 55 °C. Please consult references 3 and 6 for further information on the use of this phosphoramidite in the synthesis and purification of pseudouridine-containing oligoribonucleotides, as well as references 4,5, and 7 for studies on related versions of pseudouridine phosphoramidite. Briefly, Agris and co-workers<sup>6</sup> used standard coupling methods. Cleavage from the CPG was accomplished with ethanol saturated with anhydrous ammonia for 12-17 h at 55 °C. Desilylation was carried out using Bu<sub>4</sub>NF in THF as usual. Hall and McLaughlin<sup>3</sup> used concentrated aqueous ammonia/ethanol (3/1) for 6 h for their cleavage/deprotection. The silyl group was removed with 1 M Bu<sub>4</sub>NF in THF for 16 h at rt.

### References:

- (1) Charette, M.; Gray, M. W. *IUBMB Life* **2000**, *49*, 341-351.
- (2) Newby, M. I.; Greenbaum, N. L. *RNA*, **2001**, *7*, 833-848.
- (3) Hall, K. B.; McLaughlin, L. W. *Nucleic Acids Res.* **1992**, *20*, 1883-1889. See also reference 4 for a closely related phosphoramidite (NEtMe rather than N(*i*-Pr)<sub>2</sub>).
- (4) Gasparutto, D.; Livache, T.; Bazin, H.; Duplaa, A.-M.; Guy, A.; Khorlin, A.; Molko, D.; Roget, A.; Teoule, R. *Nucleic Acids Res.* **1992**, *20*, 5159-5166. This work describes a phosphoramidite similar to Pseudouridine CEP, but with an *N*-ethyl-*N*-methyl phosphoramidite.
- (5) Pieleś, U.; Beijer, B.; Bohmann, K.; Weston, S.; O'Loughlin, S.; Adam, V.; Sproat, B. S. *J. Chem. Soc., Perkin Trans. 1* **1994**, 3423-3429. This report describes a pseudouridine phosphoramidite bearing pivaloyloxymethyl protecting groups at N1 and N3 and an Fpmp group at the 2'-oxygen. See also the references provided therein for early non-phosphoramidite routes to pseudouridine incorporation.
- (6) Agris, P. F.; Malkiewicz, A.; Kraszewski, A.; Everett, K.; Nawrot, B.; Sochacka, E.; Jankowska, J.; Guenther, R. *Biochemie* **1995**, *77*, 125-134.
- (7) (a) Meroueh, M.; Grohar, P. J.; Qiu, J.; SantaLucia, J., Jr.; Scaringe, S. A.; Chow, C. S. *Nucleic Acids Res.* **2000**, *28*, 2075-2083. This paper describes pseudouridine incorporation using the Scaringe/Dharmacon 2'-*O*-ACE method for RNA synthesis. See also: (b) Chui, H. M.-P.; Meroueh, M.; Scaringe, S. A.; Chow, C. S. *Bioorg. Med. Chem.* **2002**, *10*, 325-332 and (c) Chui, H. M.-P.; Desaulniers, J.-P.; Scaringe, S. A.; Chow, C. S. *J. Org. Chem.* **2002**, *67*, 8847-8854.
- (8) (a) Sundaram, M.; Crain, P. F.; Davis, D. R. *J. Org. Chem.* **2000**, *65*, 5609-5614.  
(b) Bajji, A. C.; Davis, D. R. *Organic Lett.* **2000**, *2*, 3865-3868.
- (9) Burgin, A. B., Jr.; Gonzalez, C.; Matulic-Adamic, J.; Karpeisky, A. M.; Usman, N.; McSwiggen, J. A.; Beigelman, L. *Biochemistry* **1996**, *35*, 14090-14097.